Stereoselective Synthesis of cis- and trans-3,4-Dihydro-3,4,8 trihydroxynaphthalen-1(2H)-one

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A short and efficient protocol for the stereoselective synthesis of racemic trans- and cis-3,4-dihydro-3,4,8-trihydroxynaphthalen-1(2H)-one (1 and 2, resp.), is described, comprising nine and eight steps starting from commercial juglone ($=$ 5-hydroxynaphthalene-1,4-dione; 12) (Scheme 4). Furthermore, an attempt to obtain 1 and 2 via phthalide annulation as the key step (Schemes 2 and 3) and a regioselective oxidation of the intermediate 1,2,3,4-tetrahydronaphthalene-1,2,4,5-tetrols 27 and 28 with activated $MnO₂$ were carried out (Scheme 4).

Introduction. – In our investigation on phytotoxic substances produced by Ceratocystis fimbriata sp. coffea, a fungus found in the canker of the coffea tree, we have reported the isolation and structure elucidation by spectroscopic methods of *trans-* and $cis-3,4$ -dihydro-3,4,8-trihydroxynaphthalen-1(2H)-one (1 and 2, resp.) [1]. These natural polyhydroxylated α -tetralones (= 3,4-dihydronaphthalen-1(2H)-ones) are known as metabolites implicated in the branched pathway of fungal DHN-melanin biosynthesis $[2-6]$. Until now, only the natural $(-)$ -trans-isomer has been isolated from six different fungal microorganisms $[1][7-11]$. In addition, the natural *cis*-isomer known also exclusively as the $(-)$ -enantiomer has also been isolated from the mutagenic microorganism [12]. The control of the configuration of these natural products is achieved by a NADPH-dependent dehydrogenase, belonging to class $B, i.e.,$ transferring the $pro-S$ hydrogen from $C(4)$ of the nicotinamide ring to the Si face of the naphthalenol substrate [4].

Our studies on the phytotoxicity of natural samples have shown that the metabolite 1 does not seem to develop necrosis in the cells of the coffea tree [13]. Furthermore, Borgschulte and co-workers have reported that 1 is toxic in contact with the leaves of

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the poplar tree [8]. At the same time, *Bürki et al.* [14] have established that the *trans*and cis-4-hydroxyscytalones (3 and 4, resp.) generate a partial toxicity in the leaves of the plane tree. The numerous contradictions encountered in various reports concerning the beneficial or the potential toxic effect of polyhydroxylated α -tetralone derivatives in contact with different vegetable species encouraged us to reexamine the biological activity of these compounds. In relation to this reexamination, we have now developed a stereoselective synthesis of 1 and 2.

To synthesis the α -tetralones 1 and 2, we decided to use the same protocol as in the synthesis of 3,4-dihydro-2,4,8-trihydroxynaphthalen-1(2H)-one [15]. This protocol involved the ring opening of a phthalide ($=$ isobenzofuran-1(3H)-one). However, here, two alternatives could be considered. The first one involves the homologation of 7- (benzyloxy)phthalide in the presence of a Michael acceptor substituted by an alkoxy group in position 3 (Pathway A in Scheme 1). The second alternative is the ring opening of 4-(benzyloxy)phthalide with benzyl acrylate, followed by the introduction of the required additional OH group (Pathway B) or by direct decarboxylation and

subsequent oxidation ($Pathway C$). Pathway A could permit, via the direct formation of adduct 5, followed by decarboxylation, a rapid access to 1 and 2.

Results and Discussion. – The homologation reaction according to *Pathway A* was realized with 5,7-dimethoxyphthalide (6) and methyl 3-methoxy acrylate ($=$ methyl 3methoxyprop-2-enoate). However, several attempts under different conditions never yielded the desired compound but only the polysubstituted naphthalene-2-carboxylate 7 and the phthalide derivative 8 (Scheme 2).

Scheme 2. Ring Opening of 5,7-Dimethoxyphthalide (6) with Methyl 3-Methoxyacrylate (Pathway A)

a) 6 (1.0 equiv.), lithium diisopropylamide (LDA; 2.0 equiv.), MeOCH=CHCO₂Me (1.5 equiv.), THF, -40° ; 35% (18% of **7** and 17% of **8**).

Analogously to the ring opening of 7-(benzyloxy)phthalide [15], 4-(benzyloxy)phthalide (9) underwent homologation in the presence of benzyl acrylate (Pathway B or C) (Scheme 3). Unlike the 7-(benzyloxy)phthalide ring opening, the addition of the first drops of lithium diisopropylamide (LDA) to 9 gave a strongly dark-colored mixture. It is likely that the relocation of the benzylic anion at the aromatic moiety produces a strong bathochromic effect. In contrast to 7-(benzyloxy)phthalide, one equiv. of LDA was enough to convert all 4-(benzyloxy)phthalide (9) into the β -keto ester 10 (*Scheme 3*). A ¹H-NMR experiment in CDCl₃ with intermediate 10 established that the ratio between the keto forms and the enol form was 6% of the cis-isomer, 72% of enol, and 22% of the trans-isomer. The attribution of the cis- and trans-isomers and the enol form is based on the following ¹H-NMR signals: δ (H) 2.54 (ddd, J = 3.5, 4.4, 14.2, ¹H – C(3)), 4.21 (dd, J = 4.3, 12.6, H – C(2)), and 5.41 (t, J = 3.5, H – C(4)) for the *trans*-isomer, $\delta(H)$ 3.66 (*dd*, *J* = 4.3, 12.6, H – C(2)) for the *cis*-isomer, and $\delta(H)$ 2.69 $(dd, J=5.4, 17.3, H_{ax}-C(3))$ and 3.19 $(dd, J=2.6, 17.3, H_{eq}-C(3))$ for the enol form. Furthermore, the huge amount of the enol form was easily deduced from the OH signal at $\delta(H)$ 12.42. Soon, we will report the results of some phthalide annulations depending on different substitutions at the aromatic moiety; the choice of the attribution of ¹H-NMR signals to the *cis*- and *trans*-isomers or the enol form will then be discussed. In contrast to the hydrogenolysis of benzyl (benzyloxy)-1,2,3,4-tetrahydro-4-hydroxy-1 oxonaphthalene-2-carboxylate in THF, which gave isosclerone $(= 3, 4$ -dihydro-4,8dihydroxynaphthalen-1(2H)-one in 41% yield [15], the study of the direct decarboxylation of 10 under the same conditions afforded, after purification, sclerone $(= 3, 4$ dihydro-4,5-dihydroxynaphthalen-1(2H)-one; 11) in a poor yield (\lt 5%). Subsequently to this result, the study of Pathway C has been given up.

Scheme 3. Ring Opening of 4-(Benzyloxy)phthalide (9) (Pathway C)

a) 9 (1.0 equiv.), LDA (1.0 equiv.), THF, -40° . b) H₂C=CHCO₂Bn (1.5 equiv.), -10° , 1 h 30 min; 37% overall).

Like in the total synthesis of the 2,4,8-trihydroxy- α -tetralones [15], the metal enolate of 10 could be oxidized by treatment with an N-sulfonyloxaziridine and then could undergo a decarboxylation step $(Pathway B)$ or follow the reverse procedure (Pathway C). However, the risk of epimerization at $C(2)$ with exclusive formation of the *cis*-diastereoisomer during the hydrogenolysis in *Pathway B* did not encourage us to follow this method [15].

Consequently, 11 was prepared in two steps by reduction of juglone $(= 5-)$ hydroxynaphthalene-1,4-dione; 12) [16] [17] (Scheme 4). To control the stereoselectivity during the subsequent reduction of the $C=O$ group, we decided to use the di(tertbutyl)silylene protective group; thus, 11 was treated with di(tert-butyl)dichlorosilane at 80° in MeCN in the presence of Et₃N to give 13 in 83% yield. The reaction was completed in 48 h. With di(tert-butyl)silylene ditriflate ((CF_3SO_3)₂Si^tBu₂) at room temperature, the reaction occurred in 3.5 h but gave a lower yield (51%). The benzylic O-atom at C(4) of compound 13 is in the equatorial conformation (naphthalene atom numbering). In the total synthesis of the 2,4,8-trihydroxy- α -tetralones [15], a keto group was transformed to a silyl enol ether by treatment with 3 equiv. of 'BuMe₂SiOTf in the presence of Et₃N. Similarly, the sclerone derivative 13 gave silyl enol ether 14 in excellent yield with 1 equiv. of 'BuMe₂SiOTf in the presence of 1.5 equiv. of Et₃N in 1,2dichloroethane at room temperature after 20 min.

Intermediate 14 was subjected to oxidation with 3-chloroperbenzoic acid (m-CPBA) or with a catalytic amount of $OsO₄$. When the oxidation of 14 was carried out with m -CPBA, the formation of compounds 15 and 16 in favor of the derivative 15 (de 10%) was observed (*Scheme 5*). During this oxidation, only the product 15 resulted from a rearrangement of the 'BuMe₂Si group. This rearrangement allowed the easy separation of 15 and 16 by column chromatography. Thus, when the O-atom attack of m-CPBA on the C=C of the silyl enol ether moiety of 14 takes place syn to the 'Bu₂SiO group at C(4), the epoxide-ring opening leads to a pseudo-equatorial conformation of the 'BuMe₂SiO group at $C(2)$ naphthalene atom numbering as for 13. During this

Scheme 4. Total Synthesis of trans- and cis-3,4-Dihydro-3,4,8-trihydroxynaphthalen-1(2H)-ones (1 and 2, resp.)

a) SnCl₂, 2 H₂O, 4m HCl, reflux, 1 h; 75%. b) NaBH₄ (0.6 equiv.), EtOH/H₂O 99:1, r.t., 10 min; 43%. c) $t_{\rm B}$ u₂SiCl₂ (1.1 equiv.), Et₃N (5.0 equiv.), HOBt (0.1 equiv.), MeCN, 80°, 48 h; 83%. d) 'BuMe₂SiOTf' (1.0 equiv.) , Et₃N (1.5 equiv.) , 1,2-dichloroethane, r.t., 20 min; 92%. *e*) OsO₄ (2%), pyridine (cat.), $K_3[Fe(CN)_6]$ (3.0 equiv.), K_2CO_3 (3.0 equiv.), MeSO₂NH₂ (1.0 equiv.), 'BuOH/H₂O 1:1, 0°, 20 h; 56%. f) NaBH₄ (1.0 equiv.), EtOH/H₂O 99 : 1, r.t., 10 min; 82% (67% of 18 and 15% of 19), g) HF · pyridine complex (3.0 equiv.) in HF, THF/pyridine 97:3, r.t., 25 min; 96%. h) Activated MnO₂ (9.0 equiv.), MeCl/MeOH 5 : 1, r.t., 24 h; 53%. i) Buffer soln. pH 10/THF 1 : 1, r.t., 3.5 h; 53%. j) NaBH4 (1.0 equiv.), EtOH/H₂O 99 : 1, r.t., 10 min; 76%. k) HF · pyridine complex (3.0 equiv.) in HF, THF/pyridine 97:3, r.t., 2.5 h; 57%. *l*) Activated MnO₂ (9.0 equiv.), CHCl₃/MeOH 5 : 1, r.t., 24 h; 55%. *m*) HF · pyridine complex (3.0 equiv.) in HF, THF/pyridine 97 : 3, r.t., 25 min; 52%.

concerted process in which the cleavage of the Si-O bond would be simultaneous with the epoxide-ring opening, the OH group formed at $C(2)$ is able to meet the 'BuMe₂Si group and gives 15 (*Fig. 1,a*). On the other hand, when the attack of the O-atom of m-CPBA takes place *anti* to the 'Bu₂SiO group at $C(4)$, the OH group formed at $C(2)$, which is pushed in a pseudo-axial position, cannot meet the 'BuMe₂Si group during epoxide-ring opening $(Fig. 1, b)$.

Scheme 5. Oxidaton of Silyl Enol Ether 14 with m-CPBA to give Racemic 21 and 22

a) *m*-CPBA (1.0 equiv.), Cl(CH₂)₂Cl, -15° , then r.t. 1.5 h; 63% (35% of **15** and 28% of **16**). *b*) NaBH₄ (1.0 equiv.), EtOH/H₂O 10:1, 10 min, r.t.; 71% (21/22 92:8).

Fig. 1. Epoxide-ring opening to 15 and 16 (the \rightarrow represent the movement of each group formed during ring opening): a) The OH group formed at $C(2)$ is able to meet the 'BuMe₂Si group and triggers off the $^{\rm t}$ BuMe₂Si rearrangement. b) The OH group formed at C(2) and the $^{\rm t}$ BuMe₂Si group move away preventing ^tBuMe₂Si rearrangement (arbitrary atom numbering)

On oxidation with catalytic amounts of $OsO₄$, silyl enol ether 14 gave exclusively the trans compound 16 (Scheme 4). In this reaction, the control of the temperature is important to avoid formation of 17 in the basic medium. However, the ring opening of the dioxasilin of 16 in basic medium $(\rightarrow 17)$ is very useful for the control of the stereoselectivity in the subsequent reduction step. An X-ray study of intermediate 17 showed that OH-C(2) is in equatorial position, while the (hydroxysilyl)oxy group at $C(4)$ is axial (*Fig. 2*), a conformation due to an intramolecular H-bond between the Hatom of the phenolic OH group and the O-atom of the hydroxysilyl group. Furthermore, the crystal packing of 17 (Fig. 3) shows that symmetry-related molecules are linked by H-bonding involving three OH groups $(O(2), O(4))$, and $O(5)$; this forms a polymer extending in the *b* direction. On the other hand, in compound 16 , $OH-C(2)$ is pseudo-axial, while the silyloxy moiety at $C(4)$ is pseudo-equatorial. This difference of conformation between 16 and 17 was decisive during the reduction step which was

Fig. 2. View of the molecular structure of compound 17. Thermal ellipsoids drawn at the 30% probability level; arbitrary atom numbering.

Fig. 3. Packing Structure of 17

only studied with NaBH₄ in aqueous EtOH solution. Thus, reduction of 16 gave a mixture of the diastereoisomeric cis- and trans-1,2-diol 18 and 19, respectively (cis/trans 79:21), which was separated by chromatography (silica gel), whereas reduction of 17 afforded trans-1,2-diol 20 with 100% diastereoisomer excess (Scheme 4). To improve the diastereoisomer excess of the *cis*-1,2-diol (\rightarrow de 84%), compound **15** was reduced to $cis-1,2$ -diol 21 (Scheme 5).

The last steps of the total synthesis of the targets 1 and 2 were planned to consist of a series of protection and deprotection reactions followed by a regioselective oxidation of the benzylic alcohol moiety. Thus, the protection of the 1,2-diol moiety of 18 and 19 by acetylation with $Ac₂O$ in the presence of catalytic amounts of N,N-dimethylpyridin-4-amine (DMAP) in CH₂Cl₂ yielded the diacetyl derivatives 23 and 24 (Scheme 6). Subsequent deprotection of the silylene group with HF · pyridine complex provided 25 and 26. Unfortunately, the oxidation of the benzylic alcohol moiety of 25 and 26 by different methods (Dess-Martin, tetrapropylammonium perruthenate (TPAP), $MnO₂$) failed to afford the desired products and resulted in the decomposition of starting material. Finally, the targets 1 and 2 were obtained by direct deprotection of the

silylene group of 18 and 20 with HF pyridine complex in dry THF/pyridine via the tetrols 27 and 28, respectively, regioselectively oxidized at room temperature with activated MnO₂ for 20 h in MeOH/CHCl₃ 1:5 (Scheme 4).

Scheme 6. Acetylation of Compounds 18 and 19 to Give Racemic 25 and 26

a) Ac₂O (10.0 equiv.), DMAP (0.3 equiv.), CH₂Cl₂; 70 – 93%. b) HF · pyridine complex (3.0 equiv.) in HF, THF/pyridine 97 : 3, r.t., 30 min; 64 – 78%.

Conclusions. – The stereoselective total synthesis of trans- and cis-3,4-dihydro-3,4,8 trihydroxynaphthalen-1(2H)-ones (1 and 2, resp.) was elaborated in nine and eight steps, respectively, from juglone (12). The stereoselectivity was controlled by the presence of the protective di(tert-butyl)silylene group. As described earlier in the asymmetric synthesis of 3,4-dihydro-2,4,8-trihydroxy-naphthalen-1(2H)-one [15], the asymmetric Sharpless dihydroxylation of silyl enol ether 14 permitted to achieve easily the enantioselective synthesis of 1 or 2.

Currently, the total synthesis of racemic vermelone $(= 3,4$ -dihydro-3,8-dihydroxynaphthalen- $1(2H)$ -one) is described in eleven steps from 3-oxopentanedioic acid diethyl ester, and the asymmetric synthesis of $(-)$ - $(3R)$ -vermelone in five steps from the natural product $(+)$ - $(3R)$ -scytalone $(=(3R)$ -3,4-dihydro-3,6,8-trihydroxynaphthalen-1(2H)-one) [5] [6] [18] [19]. Intermediate **16** could be used to synthesize vermelone in only three steps by catalytic hydrogenolysis under pressure (5 bar) in acetone, followed by deprotection of the di(tert-butyl)silylene group and regioselective oxidation with $MnO₂$.

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Experimental Part

General. Activated MnO₂ was purchased from *Fluka* (ref. 63548). Solvents and reagents were distilled by standard procedures prior to use, and air-sensitive compounds were handled under either N_2 or Ar. Reactions were monitored by TLC. TLC: Macherey-Nagel silica gel 60 F_{254} plates; visualization by UV light and/or spraying with 5% (v/v) H₂SO₄/EtOH followed by heating. Column chromatography (CC): Macherey-Nagel silica gel 60. M.p.: Büchi-B510 apparatus. IR Spectra: Perkin-Elmer FT-IR-1720-X spectrometer; \tilde{v} in cm⁻¹. ¹H- and ¹³C-NMR Spectra: *Bruker AMX-400* spectrometer; chemicals shifts δ in ppm rel. to the residual H-atom resonances of the solvent used, J in Hz. MS: Finnigan LCQ apparatus; in m/z . Microanalyses were performed in the Mikroelementaranalytisches Laboratorium, ETH-Zürich.

Methyl 1,4-Dihydroxy-6,8-dimethoxynaphthalene-2-carboxylate (7) and Methyl 3-(4,6-Dimethoxy-3 oxoisobenzofuran-1(3H)-ylidene)propanoate (8). To 5.7-dimethoxyphthalide (200 mg, 1.03 mmol; 6) [20] in freshly distilled THF (40 ml) at -40° under Ar, commercial 2m LDA (1.03 ml, 2.06 mmol) was added dropwise with a syringe $(\rightarrow$ light-orange soln.) After 5 min stirring, methyl 3-methoxyacrylate (152 μ l, 1.54 mmol) [21] in dry THF (3 ml) was slowly added by syringe while maintaining the temp. at $-$ 40°. Then, the burgundy mixture was allowed to warm to r.t. within 2 h and quenched by the addition of 1m HCl (40 ml). The aq. layer was extracted with AcOEt (3×20 ml), the combined org. extract washed with H₂O (2×10 ml) and brine (20 ml) and dried (MgSO₄), the solvent evaporated, and the residue purified by CC (SiO₂, AcOEt/hexane 2:3): 50.2 mg (18%) of 7 and 49.3 mg (17%) of 8.

Data of 7: Pale yellow solid. R_f (AcOEt/hexane 2:3) 0.31. IR (KBr): 3400, 1655, 1631, 1611, 1261, 1220. ¹H-NMR (400 MHz, CDCl₃): 4.03 (s, MeO); 4.04 (s, MeO); 4.06 (s, MeO); 6.71 (d, J=2.4, $\rm{H-C(5)}$); 7.27 (d, J = 2.4, $\rm{H-C(7)}$); 7.30 (s, $\rm{H-C(3)}$); 8.65 (s, OH); 11.99 (s, OH). ¹³C-NMR (100 MHz, CDCl3): 51.78; 55.05; 55.75; 94.05 (C(7)); 99.13 (C(5)); 103.11; 106.42 (C(3)); 112.21; 133.58; 143.55; 156.98; 160.78; 161.20; 171.44 (C(9)). EI-MS (70 eV): 278 (M⁺), 218, 116, 91, 58.

Data of 8: White solid. M.p. $167-170^{\circ}$ (hexane/AcOEt). R_f (AcOEt/hexane 2:3) 0.22. IR (KBr): 1784, 1733, 1617, 1600. ¹H-NMR (400 MHz, CDCl₃): 3.51 $(d, J = 7.3, CH_2(2))$; 3.74 (s, MeO) ; 3.92 $(s,$ MeO); 3.96 (s, MeO); 5.74 (t, J = 7.3, H – C(3)); 6.45 (d, J = 2.8, H – C(5')); 6.68 (d, J = 2.8, H – C(7')). ¹³C-NMR (100 MHz, CDCl₃): 30.91 (C(2)); 52.09; 55.98; 56.05; 95.20 (C(5')); 99.59 (C(3)); 100.30 $(C(7'))$; 105.58; 143.19; 146.80; 159.26; 164.27; 167.03; 171.18 $(C(1))$. EI-MS (70 eV): 278 (M^+) , 219, 191, 161. Anal. calc. for C₁₄H₁₄O₆ (278.3): C 60.43, H 5.07; found: C 60.24, H 4.95.

4-(Benzyloxy)isobenzofuran-1(3H)-one $(= 4$ -(Benzyloxy)phthalide; 9). To 4-hydroxyphthalide $(3 \text{ g}, 20 \text{ mmol})$ [22] and K_2CO_3 (8.3 g, 60 mmol) in DMF (50 ml) at 0° under Ar, BnBr (3.8 g, 22 mmol) in dry DMF (10 ml) was added dropwise. The mixture was stirred at r.t. for 48 h and then quenched by addition of H₂O (50 ml) at 5°. The aq. layer was extracted with Et₂O (3×100 ml), the combined org. extract washed with brine (20 ml) and dried (MgSO₄), the solvent evaporated, and the residue purified by CC (SiO₂, AcOEt/hexane 1:1): 3.5 g (72%) of 9. White solid. M.p. 110-111^o (hexane/AcOEt). IR (KBr): 1760, 1610, 1498, 1281. ¹H-NMR (400 MHz, CDCl₃): 5.20 (s, CH₂(3)); 5.31 (s, PhCH₂); 7.18 (d, $J = 7.8, H - C(5)$; 7.36 – 7.48 (m, 6 H); 7.53 (t, $J = 8.0, H - C(6)$). ¹³C-NMR (100 MHz, CDCl₃): 68.63 (PhCH2); 70.84 (C(3)); 116.53 (C(5)); 118.03; 127.84; 127.94; 128.87; 129.20; 131.21; 135.75; 136.25; 153.79 (C(4)); 171.55 (C(1)). Anal. calc. for C_1,H_1O_3 (240.3): C 74.99, H 5.03; found: C 74.80, H 5.04.

Benzyl 5-(Benzyloxy)-1,2,3,4-tetrahydro-4-hydroxy-1-oxonaphthalene-2-carboxylate (10). To 9 $(500 \text{ mg}, 2.08 \text{ mmol})$ in freshly distilled THF (100 ml) at -40° under Ar, 2m LDA $(1.04 \text{ ml}, 2.08 \text{ mmol})$ was added dropwise with a syringe (\rightarrow dark soln.). After 5 min stirring, benzyl acrylate (439 mg, 2.70 mmol) in dry THF (5 ml) was slowly added by syringe while maintaining the temp. at -40° (\rightarrow pale yellow soln.). Then, the mixture was allowed to warm to -10° within 1 h 30 min and quenched by the addition of 1m HCl (20 ml) followed by H₂O (80 ml). The aq. layer was extracted with AcOEt (3 \times 70 ml), the combined org. extract washed with brine (25 ml) and dried (MgSO4), the solvent evaporated and the residue purified by CC (SiO₂, AcOEt/hexane 3:7): 308 mg (37%) of 10. Pale yellow oil. R_f $(ACOEthexane 3:7)$ 0.36. ¹H-NMR (400 MHz, CDCl₃): enol-10: 2.69 (dd, J = 5.4, 17.3, 1 H, CH₂(3)); $3.19 (dd, J = 2.6, 17.3, 1 \text{ H}, \text{CH}_2(3))$; $5.29 - 5.31 (m, 2 \text{ PhCH}_2\text{O}, \text{H} - \text{C}(4))$; $7.11 (dd, J = 0.9, 8.3, \text{H} - \text{C}(6))$; $7.31 - 7.48$ $(m, 12 \text{ H})$; 7.60 $(d, J = 7.8, \text{H} - \text{C}(8))$; 12.42 (s, OH) ; trans-10: 2.54 $(ddd, J = 3.5, 4.4, 14.2, 1 \text{ H})$ $CH₂(3)$); 2.65 – 2.72 (m, 1 H, CH₂(3)); 4.21 (dd, J = 4.3, 12.6, H – C(2)); 5.19 (s, PhCH₂O); 5.29 – 5.31 (m, PhCH₂O); 5.41 (t, J = 3.5, H – C(4)); 7.23 (dd, J = 1.0, 8.2, H – C(6)); 7.31 – 7.48 (m, 11 H); 7.67 (dd, J = $(0.9, 8.0, H - C(8))$; *cis-***10**: 3.66 (*dd, J* = 4.3, 12.6, H – C(2)). ¹³C-NMR (100 MHz, CDCl₃): enol-**10**: 29.28 $(C(3))$; 59.89 $(C(4))$; 66.78; 71.04; 94.37 $(C(2))$; 115.53; 117.98; 127.76; 128.18; 128.51; 128.62; 128.72; 129.05; 129.16; 129.31; 130.43; 136.24; 136.99; 155.85 (C(5)); 164.13 (C(1)); 173.03 (C(11)); trans-10: 32.95 (C(3)); 49.89 (C(2)); 60.94 (C(4)); 67.39; 71.16; 119.95; 127.38; 127.86; 127.96; 128.67; 128.89; 128.94; 129.00; 129.38; 132.28; 136.16; 136.46; 156.41 (C(5)); 170.67 (C(11)); 193.75 (C(1)). ESI-MS $(neg.): 401 ([M-H]^{-}).$

(4RS)-3,4-Dihydro-4,5-dihydroxynaphthalen-1(2H)-one (= Sclerone; 11). To β -hydrojuglone (= 2,3dihydro-5-hydroxynaphthalene-1,4-dione [16] (1.45 g, 8.24 mmol) in EtOH (180 ml) at 0° , NaBH₄ $(187 \text{ mg}, 0.494 \text{ mmol})$ in H₂O (1.8 ml) was added. The mixture was stirred for 10 min and then quenched with H₂O (100 ml). After addition of 1m HCl until pH 7 was reached, EtOH was evaporated and the aq. layer extracted with AcOEt $(3 \times 200 \text{ ml})$. Then the combined org. extract was washed with brine (150 ml) and dried $(MgSO_4)$, the solvent evaporated, and the residue purified by CC (SiO₂, AcOEt/ hexane 1:1): 632 mg (43%) of 11. White solid. R_f (AcOEt/hexane 1:1) 0.21. ¹H-NMR (400 MHz, $CDCl₃$): 1.72 (br., OH); 2.23 (m, 1 H, CH₂(3)); 2.53 – 2.61 (m, 1 H, CH₂(2), 1 H, CH₂(3)); 2.84 (dt, J = 5.0, 16.2, 1 H, CH₂(2)); 5.37 (dd, J = 4.9, 9.9, H – C(4)); 7.13 (dd, J = 1.3, 8.1, H – C(6)); 7.32 (dt, J = 0.6, 8.0, H – C(7)); 7.60 (dd, J = 1.2, 7.8, H – C(8)); 8.46 (br., OH). ¹³C-NMR (100 MHz, CDCl₃): 32.38 (C(3)); 36.01 (C(2)); 68.44 (C(4)); 119.17 (C(4a)); 122.27 (C(7)); 128.53 (C(5)); 129.50 (C(6)); 132.22 $(C(8a))$; 155.93 $(C(5))$; 196.77 $(C(1))$. ESI-MS (pos.): 179 ($[M+H]^+$).

 $(4RS)$ -4,5-{[Bis(1,1-dimethylethyl)silylene]dioxy}-3,4-dihydronaphthalen-1(2H)-one (=(3aRS)-2,2-Bis(1,1-dimethylethyl)-4,5-dihydronaphtho[1,8-de]-1,3,2-dioxasilin-6(3aH)-one; 13). To 11 (480 mg, 2.69 mmol) in dry MeCN (15 ml), HOBt ((=1-hydroxy-1H-benzotriazole; 37 mg, 0.27 mmol), Et₃N (1.88 ml, 13.5 mmol; freshly distilled from Ca H_2) and 'Bu₂SiCl₂ (626 μ l, 2.96 mmol) were added. The mixture was stirred at 80 $^{\circ}$ for 48 h. Then, the soln. was cooled to r.t. and poured into ice (25 g). The aq. layer was further extracted with Et₂O (3×30 ml), the combined org. extract washed with sat. NaHCO₃ soln. (25 ml) and brine (25 ml) and dried $(MgSO₄)$, the solvent evaporated, and the brown oily residue purified by CC (silica gel, AcOEt/hexane 1 : 9): 709 mg (83%) of 13. Pale yellow solid. Recrystallization from hexane gave a white solid. M.p. 115° (hexane). R_f (AcOEt/hexane 1:9) 0.65. IR (KBr): 2926, 2857, 1689, 1594, 1467, 1284, 1263, 829. ¹ H-NMR (400 MHz, CDCl3): 0.97 (s, Me3C); 1.17 (s, Me3C); 2.07 – 2.16 $(m, 1 \text{ H}, \text{CH}_2(3))$; 2.50 – 2.63 $(m, 1 \text{ H}, \text{CH}_2(2), 1 \text{ H}, \text{CH}_2(3))$; 2.77 – 2.84 $(m, 1 \text{ H}, \text{CH}_2(2))$; 5.30 $(dd, J =$ 4.9, 11.2, H-C(4)); 7.16 (dd, J = 1.3, 8.0, H-C(6)); 7.31 (t, J = 8.0, H-C(7)); 7.65 (dd, J = 1.3, 7.8, H-C(8)). ¹³C-NMR (100 MHz, CDCl₃): 21.56; 21.92; 27.32; 27.35; 33.17 (C(3)); 37.23 (C(2)); 69.65 (C(4)); 120.35 (C(8)); 124.92 (C(6)); 129.33 (C(7)); 132.02; 133.16; 154.17 (C(5)); 197.29 (C(1)). ESI-MS (pos.): 319 ($[M + H]^+$). Anal. calc. for C₁₈H₂₆O₃Si (318.5): C 67.88, H 8.23; found: C 68.04, H 8.17.

(1RS)-1,8-{[Bis(1,1-dimethylethyl)silylene]dioxy}-4-{[(1,1-dimethylethyl)dimethylsilyl]oxy}-1,2-dihydronaphthalene $(=(3aRS)-2,2-Bis(1,1-dimensional\text{depth})-6-[1,1-dimensional\text{depth})]$ dimethylsilyl j oxy]-3a,4 $dihydronaphtho[1,8-de]-1,3,2-dioxasilin; 14)$. To 13 (709 mg, 2.23 mmol) in 1,2-dichloroethane (20 ml) at r.t. under Ar, Et₃N (467 µl, 3.35 mmol; freshly distilled from KOH) was added by syringe. Then, at r.t., t BuMe2SiOTf (512 ml, 2.23 mmol) was added dropwise by syringe. The mixture was stirred for 25 min, then $SiO₂(5 g)$ was directly added to the soln. The solvent was evaporated and the residue purified by CC (SiO₂, AcOEt/hexane 2:98): 886 mg (92%) of 14. Colorless oil. R_f (AcOEt/hexane 2:98): 0.72. IR (KBr): 2933, 2891, 2860, 1579, 1471, 1353, 1261. ¹H-NMR (400 MHz, CDCl₃): 0.21 (s, MeSi); 0.24 (s, MeSi); 1.02 (s, Me₃C); 1.05 (s, Me₃C); 1.11 (s, Me₃C); 2.43 – 2.59 (m, CH₂(2)); 5.06 (dd, J = 2.5, 6.8, $H-C(3)$); 5.31 (dd, J=7.5, 14.0, H-C(1)); 6.84 (dd, J=1.2, 8.0, H-C(7)); 7.08 (dd, J=1.2, 7.7, $\text{H}-\text{C}(5)$); 7.16 (t, J = 7.8, $\text{H}-\text{C}(6)$). ¹³C-NMR (100 MHz, CDCl₃): -4.17 ; -4.08 ; 18.68; 21.80; 26.24; 27.23; 27.30; 32.35 (C(2)); 70.39 (C(1)); 101.81 (C(3)); 115.70 (C(5)); 119.47 (C(7)); 124.19; 128.54 $(C(6))$; 133.75; 148.04 $(C(4))$; 152.92 $(C(8))$. ESI-MS (pos.): 433 ([M+H]⁺).

(2RS,4SR)-4,5-{[Bis(1,1-dimethylethyl)silylene]dioxy}-2-{[(1,1-dimethylethyl)dimethylsilyl]oxy}- $3,4$ -dihydronaphthalen- $1(2H)$ -one $(=(3aRS,5SR)-2,2-Bis(1,1-dimethyl-v1)-5-{[(1,1-dimethyl-v1]/2,1]}$ oxy $-4,5$ -dihydronaphtho[1,8-de]-1,3,2-dioxasilin-6(3aH)-one; 15). To 14 (158 mg, 0.366 mmol) in 1,2dichloroethane (7 ml) at -15° under Ar, 70% *m*-CPBA (90 mg, 0.366 mmol) in 1,2-dichloroethane (2 ml) was added dropwise by syringe. The mixture was stirred at r.t. for 90 min. Then, the solvent was evaporated and the residue purified by CC (SiO₂, AcOEt/hexane 5:95): 57 mg (35%) of **15** and 34 mg (28%) of 16. 15: Colorless oil. R_f (AcOEt/hexane 5:95) 0.65. IR (KBr): 2934, 2861, 1699, 1595, 1468, 1281, 1263. ¹H-NMR (400 MHz, CDCl₃): 0.05 (s, MeSi); 0.18 (s, MeSi); 0.85 (s, Me₃C); 0.96 (s, Me₃C); 1.19 (s, Me₃C); 2.12 (ddd, J = 2.2, 10.2, 12.5, 1 H, CH₂(3)); 2.69 (dt, J = 4.1, 12.5, 1 H, CH₂(3)); 4.35 (dd, $J=2.1, 3.8, H-C(2))$; 5.58 (dd, $J=4.9, 10.2, H-C(4))$; 7.17 (dd, $J=1.2, 8.0, H-C(6))$; 7.33 (t, $J=8.0$, $H-C(7)$; 7.64 (dd, J = 1.2, 7.8, $H-C(8)$). ¹³C-NMR (100 MHz, CDCl₃): -4.78; -4.44; 18.52; 21.56; 25.99; 27.36; 41.47 (C(3)); 65.66 (C(4)); 72.84 (C(2)); 121.10 (C(8)); 124.80 (C(6)); 129.38 (C(7)); 130.16; 133.18; 154.11 (C(5)); 194.59 (C(1)). ESI-MS (pos.): 471 ($[M + Na]$ ⁺).

(2RS,4RS)-4,5-{[Bis(1,1-dimethylethyl)silylene]dioxy}-3,4-dihydro-2-hydroxynaphthalen-1(2H)-one $(=(3aRS,5RS)-2,2-Bis(1,1-dimethyl)-4,5-dihydro-5-hydroxynaphtho[1,8-de]-1,3,2-dioxasilin-6(3aH)-1)$ one; 16). To a soln. of K_2CO_3 (276 mg, 2.00 mmol), $K_3[Fe(CN)_6]$ (418 mg, 2.00 mmol), and pyridine (2 μ), 0.013 mmol) in H₂O (3.7 ml), 'BuOH (1 ml) and 2.5% OsO₄ soln. in 'BuOH (140 μ l, 0.013 mmol) were

added, followed by MeSO₂NH₂ (63 mg, 0.66 mmol). Then, the mixture was cooled to 0° , **14** (288 mg, 0.66 mmol) diluted in 'BuOH (2.7 ml) was added, and the mixture was stirred overnight at 0° . After quenching with Na₂SO₃ (500 mg), the mixture was stirred for 1 h at r.t., and then H₂O (10 ml) was added. The aq. layer was extracted with of AcOEt $(3 \times 25 \text{ ml})$, the combined org. extract washed with brine (20 ml) and dried (MgSO₄), the solvent evaporated, and the residue purified by CC (SiO₂, AcOEt/ hexane 1:4): 126 mg (56%) of 16. White solid. M.p. 124 $^{\circ}$ (hexane). R_f (AcOEt/hexane 2:8) 0.25. IR (KBr): 3474, 2962, 2935, 2860, 1686, 1594, 1467, 1296, 1263. ¹H-NMR (400 MHz, CDCl₃): 0.94 (s, Me₃C); 1.19 (s, Me₃C); 2.43 (ddd, J = 5.7, 7.0, 12.7, 1 H, CH₂(3)); 2.61 (ddd, J = 4.7, 7.0, 12.7, 1 H, CH₂(3)); 4.49 $(dd, J=4.7, 7.0, H-C(2))$; 5.46 $(t, J=6.3, H-C(4))$; 7.21 $(dd, J=1.1, 8.0, H-C(6))$; 7.36 $(t, J=8.0,$ $\text{H}-\text{C}(7)$); 7.58 (dd, J = 1.1, 7.7, $\text{H}-\text{C}(8)$). ¹³C-NMR (100 MHz, CDCl₃) 21.65; 21.95; 27.39; 27.45; 37.74 $(C(3))$; 65.09 $(C(4))$; 70.45 $(C(2))$; 120.39 $(C(8))$; 125.27 $(C(6))$; 129.74; 129.93 $(C(7))$; 132.37; 154.83 $(C(5))$; 197.43 $(C(1))$. ESI-MS (pos.): 357 $([M + Na]^+)$. Anal. calc. for $C_{18}H_{26}O_4Si$ (334.5): C 64.64, H 7.83; found: C 64.66, H 7.69.

(2RS,4RS)-4-{[Bis(1,1-dimethylethyl)hydroxysilyl]oxy}-3,4-dihydro-2,5-dihydroxynaphthalen-1(2H) one (17) . A soln. of 16 (62 mg, 0.186 mmol) in buffer soln. pH 10/THF 1:1 (10 ml) was stirred at r.t. for 90 min and then neutralized with 2m HCl (\rightarrow pH 7). The aq. layer was extracted with AcOEt (3 \times 20 ml), the combined org. extract washed with brine (20 ml) and dried $(MgSO₄)$, the solvent evaporated, and the residue purified by CC (SiO₂, AcOEt/hexane 1:4): 35 mg (53%) of 17. White solid. R_f (AcOEt/hexane 2 : 8) 0.15. IR (KBr): 3435, 2935, 2860, 1694, 1062. ¹ H-NMR (400 MHz, CDCl3): 0.85 (s, Me); 1.07 (s, Me, $Me₃C$); 1.16 (s, Me); 2.23 (dt, J = 2.8, 13.0, 1 H, CH₂(3)); 2.83 (ddd, J = 3.3, 5.2, 13.0, 1 H, CH₂(3)); 5.07 $(dd, J=5.1, 12.7, H-C(2))$; 5.52 $(t, J=2.9, H-C(4))$; 7.21 $(dd, J=1.1, 7.7, H-C(6))$; 7.38 $(t, J=8.0,$ $\text{H}-\text{C}(7)$); 7.71 (dd, J = 1.1, 7.7, $\text{H}-\text{C}(8)$). ¹³C-NMR (100 MHz, CDCl₃): 20.19; 20.51; 27.10; 27.63; 27,67; 40.39 (C(3)); 63.27 (C(4)); 69.21 (C(2)); 120.65 (C(8)); 124.80 (C(6)); 130.46; 130.62 (C(7)); 130.69; 154.29 (C(5)); 200.18 (C(1)). ESI-MS (neg.): 351 ($[M-H]$ ⁻).

X-Ray Crystal Structure of 17. X-Ray crystal data for $C_{18}H_{28}O_5Si$ (M_r 352.51): Orthorhombic space group *Pbca*, $D_c = 1.229 \text{ g} \cdot \text{cm}^{-3}$, $Z = 8$; $a = 11.640$, $b = 14.381$, $c = 22.761 \text{ Å}$, $\alpha = \beta = \gamma = 90^{\circ}$, $V =$ 3809.8 \AA ³; Mo K_a radiation, $\lambda = 0.71073 \AA$, 2.42 $\lt \theta$ $\lt 25.89^\circ$; number of reflections measured 27881, number of independent reflections 3666, number of observed reflections 3056 ($I > 2\sigma(I)$), T 153 K. Suitable crystals of 17 were grown from benzene as pale yellow blocks. Intensity data were collected at 153 K with a Stoe-Image-Plate-Diffraction system [23] and $M \alpha K_{\alpha}$ graphite-monochromated radiation. Image-plate distance 70 mm, ϕ oscillation scans 0-198°, step $\Delta \phi = 1.0^{\circ}$, 2 θ range 3.27-52.1°, d_{max} $d_{\text{min}} = 12.45 - 0.81$ Å. The structure was solved by direct methods with the program SHELXS-97 [24]. The refinement and all further calculations were carried out with SHELXL-97 [25]. The H-atoms were located from Fourier difference maps and refined isotropically. The non-H-atoms were refined anisotropically by means of weighted full-matrix least-squares on $F²$. All-data refinement of 319 parameters based on F^2 converged at $R(F) = 0.0431$ and $wR(F^2) = 0.0955$. The molecular structure and crystallographic numbering scheme are illustrated in the PLATON drawing [26] (Fig. 2) CCDC-602497 contains the supplementary crystallographic data. These data can be obtained free of charge via http:// www.ccdc.cam.ac.uk/data_request.cif.

(1RS,2SR,4SR)- and (1RS,2RS,4RS)-4,5-{[Bis(1,1-dimethylethyl)silylene]dioxy}-1,2,3,4-tetrahydronaphthalene-1,2-diol ($=(3aRS, 5RS, 6SR)$ - and ($3aRS, 5RS, 6RS$)-2,2-Bis(1,1-dimethylethyl)-3a,4,5,6-tetrahydronaphtho[1,8-de]-1,3,2-dioxasilin-5,6-diol, resp.; **18** and **19**). To a soln. of **16** (277 mg, 0.829 mmol) in EtOH (20 ml) at r.t., $NabH_4$ (32 mg, 0.829 mmol) in H_2O (4 ml) was added. The mixture was stirred for 10 min and then H₂O (20 ml) was added. After neutralization of the soln. (\rightarrow pH 7) with 1m HCl, EtOH was evaporated and the aq. layer extracted with AcOEt $(3 \times 40 \text{ ml})$. The combined org. extract was washed with brine (20 ml) and dried $(MgSO₄)$, the solvent evaporated, and the residue purified by CC $(SiO₂, AcOEt/hexane 3:7)$: 186 mg (67%) of 18 and 43 mg (15%) of 19.

Data of 18: White solid. M.p. 120° (hexane/AcOEt). R_f (AcOEt/hexane 3:7) 0.30. IR (KBr): 3390, 2934, 2860, 1602, 1584, 1473, 1455, 1266, 968. ¹ H-NMR (400 MHz, CDCl3): 0.94 (s, Me3C); 1.15 (s, Me3C); 1.81 (ddd, J = 1.8, 10.2, 12.0, 1 H, CH₂(3)); 2.59 (dt, J = 5.6, 12.0, 1 H, CH₂(3)); 3.00 (br., OH); 4.17 – 4.20 $(m, H-C(2))$; 4.64 $(d, J = 3.4, H-C(1))$; 5.34 $(dd, J = 5.9, 10.2, H-C(4))$; 6.85 $(d, J = 7.9, H-C(6))$; 7.10 $(d, J = 7.7, H - C(8))$, 7.22 $(t, J = 7.8, H - C(7))$. ¹³C-NMR (100 MHz, CDCl₃): 21.71; 21.87; 27.51; 27.59;

37.12 (C(3)); 66.27 (C(4)); 69.42 (C(2)); 70.30 (C(1)); 118.47 (C(6)); 120.37 (C(8)); 127.19; 129.51 $(C(7))$; 136.73; 154.41 $(C(5))$. ESI-MS (pos.): 359 ([M+Na]⁺).

Data of **19**: White solid. R_f (AcOEt/hexane 3:7) 0.23. ¹H-NMR (400 MHz, CDCl₃ + D₂O): 0.91 (s, Me_3C); 1.15 (s, Me_3C); 2.25 (t, J = 5.9, CH₂(3)); 3.98 (q, J = 5.9, H – C(2)); 4.44 (d, J = 6.0, H – C(1)); 5.23 $(t, J=6.7, H-C(4))$; 6.89 (dd, $J=0.5, 7.9, H-C(6)$); 7.07 (d, $J=7.7, H-C(8)$); 7.24 (t, $J=7.9, H-C(7)$). ¹³C-NMR (100 MHz, CDCl₃): 21.65; 21.92; 27.49; 27.52; 35.66 (C(3)); 66.31 (C(4)); 70.75 (C(2)); 72.96 $(C(1))$; 118.94 $(C(6))$; 120.27 $(C(8))$; 126.59; 129.72 $(C(7))$; 136.75; 154.58 $(C(5))$. ESI-MS (pos.): 359 $([M + Na]^{+}).$

(1RS,2RS,4RS)-4-{[Bis(1,1-dimethylethyl)hydroxysilyl]oxy}-1,2,3,4-tetrahydronaphthalene-1,2,5-tri ol (20). As described for 18/19, with 17 (41 mg, 0.116 mmol) in EtOH (3 ml) and NaBH₄ soln. (600 µ, $c =$ 9.3 g 1^{-1} , 0.14 mmol) in H₂O: 32 mg (76%) of **20**. White solid. R_f (AcOEt/hexane 1:1) 0.22. IR (KBr): 3412, 2932, 2858, 1589, 1471, 1053. ¹H-NMR (400 MHz, CD₃COCD₃ + D₂O): 0.84 (s, Me); 0.99 (s, Me, $Me₃C$); 1.06 (s, Me); 1.83 (ddd, J = 3.2, 11.8, 13.1, 1 H, CH₂(3)); 2.37 (dt, J = 3.4, 13.1, 1 H, CH₂(3)); 4.23 $(ddd, J = 3.3, 8.4, 11.7, H - C(2))$; 4.34 $(d, J = 8.5, H - C(1))$; 5.34 $(t, J = 3.2, H - C(4))$; 6.76 $(ddd, J = 0.7,$ 1.2, 7.5, H-C(6)); 7.12–7.17 (m, H-C(7), H-C(8)). ¹³C-NMR (100 MHz, CD₃COCD₃): 20.10; 20.45; 20.99; 27.11; 27.38; 27.51; 39.44 (C(3)); 64.75 (C(4)); 68.82 (C(2)); 75.80 (C(1)); 115.33 (C(6)); 119.61; 124.83; 129.30; 140.09; 154.75 (C(5)). ESI-MS (neg.): 353 ($[M-H]$ ⁻).

(1RS,2SR,4RS)- and (1RS,2RS,4SR)-4,5-{[Bis(1,1-dimethylethyl)silylene]dioxy}-2-{[(1,1-dimethyl $ethyl] dimethylsilyl] oxy] -1,2,3,4-tetrahydronaphthalen-1-ol (= (3aRS,5SR,6SR) - and (3aRS,5SR,6SR) -$ 2,2-Bis(1,1-dimethylethyl)-5-{[(1,1-dimethylethyl)dimethylsilyl]oxy}-3a,4,5,6-tetrahydronaphtho[1,8-de]- 1,3,2-dioxasilin-6-ol; 21 and 22, resp.). As described for 18/19, with 15 (23 mg, 0.0513 mmol) in EtOH (1 ml) and NaBH₄ soln. $(100 \mu, c = 19.5 \text{ g l}^{-1}, 0.0513 \text{ mmol})$ in H₂O: 17 mg (71%) of 21/22 (ratio 92:8). Colorless oil mixture.

Data of 21: Colorless oil. Hexane/AcOEt. R_f (AcOEt/hexane 3:7) 0.36. ¹H-NMR (400 MHz, CDCl₃): 0.16 (s, Me); 0.19 (s, Me); 0.88 (s, Me₃C); 0.95 (s, Me₃C); 1.15 (s, Me₃C); 1.85 (ddd, J = 1.6, 10.3, 13.5, 1 H, CH₂(3)); 2.45 (d, J = 10.8, OH); 2.53 (dt, J = 5.6, 13.5, 1 H, CH₂(3)); 4.36 (ddd, J = 1.6, 3.8, 5.5, $H-C(2)$; 4.61 (dd, J = 3.8, 10.8, H - C(1)); 5.34 (dd, J = 5.6, 10.3, H - C(4)); 6.84 (d, J = 7.9, H - C(6)); 7.20 – 7.26 $(m, H - C(7), H - C(8))$. ¹³C-NMR (100 MHz, CDCl₃): -4.83 ; -4.49 ; 14.54; 18.50; 21.65; 26.14; 27.46; 27.51; 38.51 (C(3)); 66.49 (C(4)); 70.20 (C(1)); 70.65 (C(2)); 118.01 (C(6)); 121.00; 127.02; 129.21; 137.99; 153.89 (C(5)). ESI-MS (neg.): 450 ($[M-H]$ ⁻).

Data of 22: Colorless oil. Hexane/AcOEt. R_f (AcOEt/hexane 3:7) 0.36. ¹H-NMR (400 MHz, CDCl₃): 0.23 (s, Me); 0.26 (s, Me); 0.94 (s, Me₃C); 1.00 (s, Me₃C); 1.15 (s, Me₃C); 1.81 – 1.89 (m, 1 H, $CH₂(3)$); 2.70 (dt, J = 5.6, 13.5, 1 H, CH₂(3)); 4.20–4.24 (m, H–C(2)); 4.80 (d, J = 5.6, H–C(1)); 5.40 $(dd, J=5.6, 10.3, H-C(4))$; 6.84 $(d, J=7.9, H-C(6))$; 6.99 $(d, J=7.8, H-C(8))$; 7.21 – 7.24 $(m, H-C(7))$. 13 C-NMR (100 MHz, CDCl₃): -4.28 ; -3.84 ; 21.68; 21.87; 21.97; 26.23; 27.51; 27.61; 36.77 (C(3)); 66.37 $(C(4))$; 69.77 $(C(2))$; 71.51 $(C(1))$; 118.21 $(C(6))$; 120.50 $(C(8))$; 127.27; 129.13 $(C(7))$; 136.84; 154.35 $(C(5))$. ESI-MS (neg.): 450 ([*M* – H]⁻).

(1RS,2SR,4SR)-4,5-{[Bis(1,1-dimethylethyl)silylene]dioxy}-1,2,3,4-tetrahydronaphthalene-1,2-diyl $Diaceate (= (3aRS,5RS,6SR)-2,2-Bis(1,1-dimethyl)-3a,4,5,6-tetrahydronaphtho[1,8-de]-1,3,2-dioxa-1)$ silin-5,6-diol Diacetate; 23). To a soln. of 18 (92 mg, 0.274 mmol) in dry CH₂Cl₂ (5 ml) at r.t. under Ar, DMAP (10 mg, 0.0822 mmol) and then dry Ac $O(259 \text{ ul}, 0.274 \text{ mmol})$ were added by syringe. The mixture was stirred for 48 h and then quenched with $H_2O(5 \text{ ml})$. The aq. layer was extracted with CH_2Cl_2 $(3 \times 10 \text{ ml})$, the combined org. extract washed with brine (20 ml) and dried (MgSO₄), the solvent evaporated, and the residue purified by CC (SiO₂, AcOEt/hexane 3:7): 107 mg (93%) of 23. White solid. M.p. 116 – 117° (hexane/AcOEt). R_f (AcOEt/hexane 3:7) 0.44. IR (KBr): 2934, 2858, 1749, 1586, 1268, 1249. ¹ H-NMR (400 MHz, CDCl3): 0.92 (s, Me3C); 1.17 (s, Me3C); 2.05 (s, MeCOO); 2.03 – 2.10 (m, $1 \text{ H, CH}_2(3)$); 2.14 (s, MeCOO); 2.62 (dt, J = 6.3, 12.8, 1 H, CH₂(3)); 5.31 (dd, J = 6.4, 8.9, H – C(4)); 5.61 $(ddd, J=2.8, 6.2, 8.9, H-C(2))$; 6.10 $(d, J=3.5, H-C(1))$; 6.84 $(d, J=7.7, H-C(6))$; 6.91 $(d, J=8.0, 1.4)$ $\text{H}-\text{C}(8)$); 7.23 (t, J = 8.0, $\text{H}-\text{C}(7)$). ¹³C-NMR (100 MHz, CDCl₃): 21.36; 21.43; 21.71; 21.90; 27.46; 27.54; 34.58 (C(3)); 66.37 (C(4)); 68.96 (C(2)); 70.05 (C(1)); 119.16 (C(8)); 120.05 (C(6)); 127.29 (C(7)); 129.59; 132.64; 154.73 (C(5)); 170.95; 171.05. ESI-MS (pos.): 443 ($[M + Na]^+$). Anal. calc. for $C_2H_{32}O_6Si$ (420.6): C 62.83, H 7.67; found: C 62.85, H 7.55.

(1RS,2RS,4RS)-4,5-{[Bis(1,1-dimethylethyl)silylene]dioxy}-1,2,3,4-tetrahydronaphthalene-1,2-diyl $Diaceate (= \frac{3aRS5RS6RS}{2.2-Bis(1.1-dimethylethvl)-3a.4.5.6-tetrahvdronaphtho[1.8-del-1.3.2-dioxa-1]$ silin-5,6-diol Diacetate; 24). As described for 23, with 19 (14 mg, 0.0417 mmol) in dry CH₂Cl₂ (2 ml), DMAP (1.5 mg, 0.0125 mmol), and Ac₂O (39 μ l, 0.417 mmol): 12 mg (70%) of **24.** White solid. R_f (AcOEt/hexane 3:7) 0.44. IR (KBr): 2963, 2934, 2860, 1747, 1602, 1584, 1474, 1368, 1247, 1227. ¹H-NMR $(400 \text{ MHz}, \text{CDC1}_3): 0.96 \text{ (s, Me}_3\text{C}); 1.16 \text{ (s, Me}_3\text{C}); 2.05 \text{ (s, MeCOO)}; 2.11 \text{ (s, MeCOO)}; 2.12-2.18 \text{ (m, m, s)}$ $1 \text{ H, CH}_2(3)$; 2.51 (ddt, J = 1.2, 4.3, 13.9, 1 H, CH₂(3)); 5.18 (dd, J = 5.9, 10.6, H – C(4)); 5.23 (ddd, J = $2.8, 5.8, 7.0, H-C(2))$; 5.87 (d, J = 2.7, H - C(1)); 6.93 – 6.95 (m, H - C(6), H - C(7)); 7.22 (dt, J = 0.6, 8.0, H-C(8)). ¹³C-NMR (100 MHz, CDCl₃) 21.45; 21.60; 21.64; 21.92; 27.42; 27.50; 32.51 (C(3)); 65.97 $(C(4))$; 68.80 $(C(1))$; 70.44 $(C(2))$; 119.71; 123.02; 127.95; 129.58 $(C(7))$; 132.34; 154.44 $(C(5))$; 170.25; 170.34. ESI-MS (pos.): 443 ($[M + Na]^+$).

 $(1RS, 2SR, 4SR)$ -1,2,3,4-Tetrahydro-4,5-dihydroxynaphthalene-1,2-diyl Diacetate (=(1RS,2SR,6SR)-1,2,3,4-Tetrahydronaphthalene-1,2,4,5-tetrol 1,2-Diacetate; **25**). To a soln. of **23** (90 mg, 0.214 mmol) in dry THF (5 ml) in a plastic flask at r.t., and dry pyridine (323 μ , 2.97 mmol) and then 70% HF/pyridine (161 mg, 4.27 mmol) were added by syringe. The mixture was stirred for 30 min, then AcOEt (15 ml) was added. The mixture was washed with brine (10 ml) and dried $(MgSO₄)$, the solvent evaporated, and the residue purified by CC (SiO₂, AcOEt/hexane 1:1): 47 mg (78%) of 25. White solid. M.p. 141 \degree (hexane/ AcOEt). R_f (AcOEt/hexane 1:1) 0.22. IR (KBr): 3481, 3242, 1734, 1719, 1595, 1471, 1368, 1260, 1229. $1\,\text{H-NMR}$ (400 MHz, CD₃COCD₃): 1.99 (s, MeCOO); 2.04 (s, MeCOO); 2.06–2.10 (m, 1 H, CH₂(3)); 2.45 (ddd, $J = 5.1, 10.8, 13.4, 1$ H, CH₂(3)); 3.84 (br., OH); 5.26 (t, $J = 4.7, H - C(4)$); 5.53 (dt, $J = 3.3, 10.8$, $\rm{H-C(2)}$); 6.13 (d, J = 3.4, $\rm{H-C(1)}$); 6.78 (d, J = 7.7, $\rm{H-C(6)}$); 6.88 (dd, J = 1.1, 8.0, $\rm{H-C(8)}$); 7.19 (t, J = 7.8, H-C(7)). ¹³C-NMR (100 MHz, CD₃COCD₃) 20.44; 20.46; 32.48 (C(3)); 63.58 (C(4)); 67.91 (C(2)); 69.07 (C(1)); 116.10 (C(8)); 120.83 (C(6)); 125.33; 129.64 (C(7)); 134.15; 156.27 (C(5)); 170.11; 170.27. ESI-MS (neg.): 279 ($[M-H]$ ⁻). Anal. calc. for C₁₄H₁₆O₆ (280.3): C 60.00, H 5.75; found: C 60.14, H 5.91.

(1RS,2RS,4RS)-2-Acetoxy-4,5-dihydroxy-1,2,3,4-tetrahydronaphthalene-1,2-diyl Diacetate $(=(1RS, 2RS, 3RS) - 1, 2, 3, 4-Tetrahydronaphthalene-1, 2, 4, 5-tetrol 1, 2-Diacetate; 26)$. As described for 25, with 24 (15 mg, 0.0286 mmol) in dry THF (2 ml), dry pyridine (76 μ , 0.698 mmol), and 70% HF/pyridine $(25 \text{ mg}, 0.662 \text{ mmol})$: 5 mg (64%) of 26. White solid. R_f (AcOEt/hexane 1:1) 0.24. ¹H-NMR (400 MHz, CD_3COCD_3 : 2.01 (s, MeCOO); 2.10 (s, MeCOO); 2.26 – 2.29 (m, CH₂(3)); 4.74 (br., OH); 5.19 – 5.22 $(m, H-C(4)); 5.37 - 5.42$ $(m, H-C(2)); 5.92$ $(d, J = 6.4, H-C(1)); 6.75$ $(d, J = 7.7, H-C(6)); 6.83$ $(dd, J =$ 1.1, 8.1, H-C(8)); 7.18 (t, J=7.8, H-C(7)); 8.86 (br., OH). ¹³C-NMR (100 MHz, CD₃COCD₃): 20.43; 20.49; 33.79 (C(3)); 63.77 (C(4)); 69.58 (C(2)); 71.11 (C(1)); 115.75 (C(8)); 119.94 (C(6)); 125.23; 129.48 $(C(7))$; 134.53; 156.46 $(C(5))$; 169.77; 170.25. ESI-MS (neg.): 279 ([M-H]⁻).

(1RS,2SR,4SR)-1,2,3,4-Tetrahydronaphthalene-1,2,4,5-tetrol (27). To a soln. of 18 (103 mg, 0.307 mmol) in dry THF (10 ml) in a plastic flask at r.t., dry pyridine (465 μ l, 4.27 mmol) and then 70% HF/pyridine (231 mg, 6.11 mmol) were added by syringe. The mixture was stirred for 30 min and then AcOEt (15 ml) was added. The mixture was poured over $SiO₂$ (1.5 g), the solvent evaporated, and the residue purified by CC (SiO₂, AcOEt): 58 mg (96%) of 27. White solid. R_f (AcOEt) 0.24. ¹H-NMR $(400 \text{ MHz}, \text{CD}_3\text{OD})$: 1.95 $(ddt, J = 1.0, 3.5, 13.4, 1 \text{ H}, \text{CH}_2(3))$; 2.30 $(ddd, J = 4.8, 11.1, 13.4, 1 \text{ H}, \text{CH}_2(3))$; 4.24 $(dt, J = 3.3, 11.1, H - C(2))$; 4.61 $(d, J = 3.5, H - C(1))$; 5.20 $(t, J = 4.2, H - C(4))$; 6.78 $(dd, J = 1.1, 8.0$, $H-C(8)$); 6.91 (dd, J = 0.6, 7.6, $H-C(6)$); 7.17 (t, J = 7.8, $H-C(7)$). ¹³C-NMR (100 MHz, CD₃OD): 34.26 $(C(3))$; 63.91 $(C(4))$; 66.67 $(C(2))$; 70.09 $(C(1))$; 114.74 $(C(8))$; 121.16 $(C(6))$; 124.21; 129.16 $(C(7))$; 138.58; 156.08 (C(5)). ESI-MS (neg.): 195 ($[M-H]$ ⁻).

(1RS,2RS,4RS)-1,2,3,4-Tetrahydronaphthalene-1,2,4,5-tetrol (28). As described for 27, with 19 (20 mg, 0.0594 mmol) in dry THF (5 ml) dry pyridine (90 μ l, 0.828 mmol), and 70% HF/pyridine $(45 \text{ mg}, 1.19 \text{ mmol})$: 6 mg (52%) of 28. White solid. As described for 27, with 20 (14 mg, 0.0395 mmol) in dry THF (1.4 ml), dry pyridine (63 μ , 0.578 mmol), and 70% HF/pyridine (32 mg, 8.48 mmol) but for 2 h 30 min: 4.4 mg (57%) of 28. White solid. R_f (AcOEt) 0.15. ¹H-NMR (400 MHz, CD₃OD): 1.92 (*ddd, J* = $4.5, 11.0, 13.5, 1 \text{ H}, \text{CH}_2(3)$; $2.22 \text{ (dt, J = 3.6, 13.5, 1 H, CH₂(3))}$; $4.05 \text{ (ddd, J = 3.3, 7.8, 11.0, H-C(2))}$; 4.33 $(d, J = 7.7, H - C(1))$; 5.15 $(t, J = 4.1, H - C(4))$; 6.73 $(dd, J = 0.7, 1.1, 7.9, H - C(8))$; 7.06 $(dt, J = 0.9,$ 7.8, H $-C(6)$); 7.16 (t, J = 7.9, H $-C(7)$). ¹³C-NMR (100 MHz, CD₃OD): 37.25 (C(3)); 63.48 (C(4)); 68.96 $(C(2))$; 74.44 $(C(1))$; 113.90 $(C(8))$; 118.63 $(C(6))$; 124.04; 129.10 $(C(7))$; 139.71; 155.82 $(C(5))$. ESI-MS $(neg.): 195 ([M-H]^{-}).$

cis-(3RS,4SR)-3,4-Dihydro-3,4,8-trihydroxynaphthalen-1(2H)-one (2). Under stirring, 27 (31 mg, 0.158 mmol) was dissolved in MeOH (1.7 ml) , and then CHCl $_3$ (8.6 ml) was added. At r.t., activated MnO₂ (151 mg, ca. 1.56 mmol; *Fluka* ref. 63548) was added by spatula. The mixture was stirred overnight and then filtered over $SiO₂$. The $SiO₂$ was washed with AcOEt and then the solvent evaporated: 16.3 mg (53%) of 2. Pale yellow solid. R_f (AcOEt) 0.33. ¹H-NMR (400 MHz, CD₃OD): 2.88 (dd, J = 3.9, 17.4, 1 H, $CH₂(2)$); 2.99 (dd, J = 7.2, 17.4, 1 H, CH₂(2)); 4.30 (ddd, J = 2.8, 3.9, 6.9, H – C(3)); 4.86 (d, J = 2.8, $H-C(4)$); 6.88 (ddd, $J=0.5, 1.0, 8.4, H-C(5)$); 7.12 (dt, $J=1.0, 7.5, H-C(7)$); 7.54 (dd, $J=7.5, 8.4$, $\text{H}-\text{C}(6)$). ¹³C-NMR (100 MHz, CD₃OD): 42.90 (C(2)); 69.45 (C(3)); 70.29 (C(4)); 115.65 (C(9)); 116.82 $(C(5))$; 119.02 $(C(7))$; 136.93 $(C(6))$; 144.65 $(C(10))$; 162.22 $(C(5))$; 203.72 $(C(1))$. ESI-MS (neg.): 193 $([M - H]^{-}).$

trans-(3RS,4RS)-3,4-Dihydro-3,4,8-trihydroxynaphthalen-1(2H)-one (1). As described for 2, with 28 $(6 \text{ mg}, 0.0306 \text{ mmol})$, MeOH (0.3 ml) , CHCl₃ (1.7 ml) , and activated MnO₂ $(30 \text{ mg}, ca, 0.3 \text{ mmol})$; 3.3 mg (55%) of 1. Pale yellow solid. R_f (AcOEt) 0.33. ¹H-NMR (400 MHz, CD₃OD): 2.72 (*dd*, $J = 8.0$, 17.2, 1 H, CH₂(2)); 3.11 (dd, J = 4.0, 17.2, 1 H, CH₂(2)); 4.05 – 4.11 (m, H – C(3)); 4.63 (d, J = 7.0, $H-C(4)$; 6.88 (dd, $J=0.6, 8.6, H-C(5)$); 7.14 (dt, $J=0.6, 7.4, H-C(7)$); 7.56 (dd, $J=7.4, 8.4, H-C(6)$). ESI-MS (neg.): 193 ($[M-H]$ ⁻).

REFERENCES

- [1] G. Gremaud, R. Tabacchi, Phytochemistry 1996, 42, 1547.
- [2] M. H. Wheeler, R. D. Stipanovic, Arch. Microbiol. 1985, 142, 234.
- [3] G. S. Basarab, J. J. Steffens, Z. Wawrzak, R. S. Schwartz, T. Lundqvist, D. B. Jordan, Biochemistry 1999, 38, 6012.
- [4] J. E. Thompson, G. S. Basarab, A. Andersson, Y. Lindqvist, D. B. Jordan, Biochemistry 1997, 36, 1852.
- [5] F. Viviani, M. Gaudry, A. Marquet, New J. Chem. 1992, 16, 81.
- [6] F. Viviani, M. Gaudry, A. Marquet, J. Chem. Soc., Perkin Trans. 1 1990, 1255.
- [7] S. Iwasaki, H. Muro, S. Nozoe, S. Okuda, Tetrahedron Lett. 1972, 13, 13.
- [8] K. Borgschulte, S. Rebuffat, W. Trowitszch-Kienast, D. Schomburg, J. Pinon, B. Bodo, Tetrahedron 1991, 47, 8351.
- [9] P. Venkatasubbaiah, W. S. Chilton, Mycopathologia 1992, 120, 33.
- [10] H. Tabuchi, A. Tajimi, A. Ichihara, Biosci., Biotechnol., Biochem. 1994, 58, 1956.
- [11] B. Nicolet, R. Tabacchi, 'Modern Fungicides and Antifungal Compounds II', 12th International Reinhardsbrunn Symposium, 1999, p. 469.
- [12] R. D. Stipanovic, A. A. Bell, Mycologia 1977, 69, 164.
- [13] G. Gremaud, Ph.D. Thesis, University of Neuchâtel, 1996.
- [14] N. Bürki, A. Michel, R. Tabacchi, Phytopathol. Mediterr. 2003, 42, 191.
- [15] E. Couché, A. Fkyerat, R. Tabacchi, *Helv. Chim. Acta* 2003, 86, 210.
- [16] R. H. Thomson, *J. Chem. Soc.* **1950**, 1737.
- [17] A. I. Gurevich, M. N. Kosolov, L. N. Nametkina, Bull. Acad. Sci. USSR Div. Chem. Sci. 1968, 1194.
- [18] B. W. Bycroft, M. M. Cashyap, T. K. Leung, J. Chem. Soc., Chem. Commun. 1974, 443.
- [19] F. Viviani, Ph.D. Thesis, University of Paris VI, 1990.
- [20] P. D. Noire, R. W. Franck, Synthesis 1980, 882.
- [21] B. Fölisch, W. Giering, Synthesis 1980, 231.
- [22] M. A. Buehler, T. A. Powers, J. G. Michels, J. Am. Chem. Soc. 1944, 66, 417.
- [23] IPDS Software, Stoe & Cie GmbH, Darmstadt, Germany, 2000.
- [24] G. M. Sheldrick, SHELXS97, Program for Crystal Structure Determination, Acta Crystallogr., Sect. A 1990, 46, 467.
- [25] G. M. Sheldrick, SHELXL97, Program for the Refinement of Crystal Structures, Universität Göttingen, Göttingen, Germany, 1997.
- [26] A. L. Spek, PLATON/PLUTON (version Jan. 1999), Acta Crystallogr., Sect. A 1990, 46 (Suppl.), $c³4$

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